

EVALUATING THE POTENTIAL EFFECT OF ETHANOL TREATMENT ON WHEY PROTEINS DIGESTIBILITY



Charikleia Kyrkou¹, Konstantina Tenzidou¹, Asterios Stamkopoulos¹, Foteini Tsakoumaki¹, Garoufalia Charitou¹, Thomas Moschakis¹, Costas G. Biliaderis¹, Alexandra-Maria Michaelidou¹

¹Department of Food Science and Technology, School of Agriculture, Faculty of Agriculture, Forestry and Natural Environment, Aristotle University of Thessaloniki, Thessaloniki 541 24, Greece

Introduction

Whey proteins (WP), although exhibiting high nutritional and functional attributes, demonstrate significant resistance to hydrolysis phenomena due to their globular structure [1; 2]. Studies have indicated that, in comparison with the traditional heat treatment, ethanol can irreversibly and more effectively denature proteins and change their secondary structures [3; 4]. However, to the best of our knowledge, data regarding the effect of ethanol on WP digestibility is scarce.

Objectives

This study focuses on the comparison of the degradation pattern and evaluation of differences in digestibility of WP under four different conditions, namely: in native WP isolates aqueous solution (**N-WPI**), in water-ethanol WP solutions with different ethanol concentrations (i.e., 10% and 50% w/v, **E-10 WPI**, and **E-50 WPI**, respectively) and in WP aqueous solution obtained after heating at 90°C for 10 min (**H-WPI**).

Method/Design

The protein content was measured with the Kjeldahl method according to ISO 8968-3:2007/IDF 20-3: 2007 [5]. To simulate the physiological digestion process (i.e., the oral (**OP**), gastric (**GP**), and intestinal phases (**IP**)), all substrates were subjected to the INFOGEST static *in vitro* digestion protocol [6]. The enzyme activities (pepsin and trypsin) were quantified prior to the implementation of the INFOGEST protocol as proposed by Minekus *et al.* (2014) [7]. A discontinuous polyacrylamide gel electrophoresis (disc-PAGE) was initially applied on the samples obtained from the *in vitro* digestion (4 treatments x 3 phases) [8]. Furthermore, Sodium Dodecyl Sulphate PAGE (SDS-PAGE) was carried out following the method of Laemmli (1970) [9] with some modifications. A Molecular Weight Marker (MwM) composed of a mixture of standard proteins of known molecular weight (from 14.2 kDa to 66.3 kDa) was also run in parallel on the same gel. All experiments were conducted in triplicates and on a protein-equivalent basis.

Results

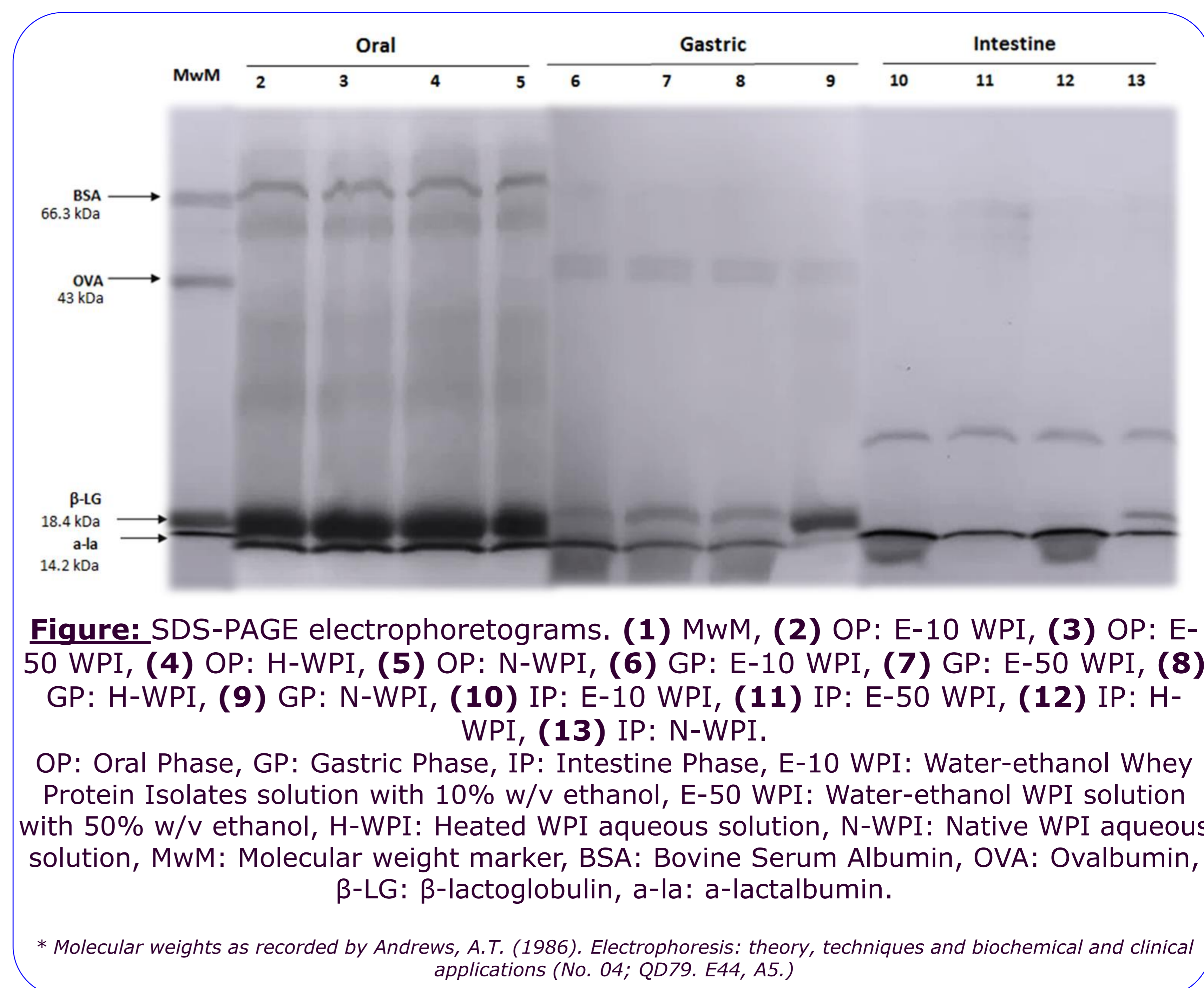
The mean protein content of undigested substrates, as well as OP, GP, and IP samples was 3.57% (± 0.01), 1.84% (± 0.07), 1.00 (± 0.04) and, 0.55% (± 0.02), respectively.

Results of disc-PAGE showed that:

- The OP samples gave, in all 4 substrates, bands with the same electrophoretic mobility (EM) as the protein zones corresponding to α -lactalbumin (α -la) and β -lactoglobulin (β -LG), ran on the same gel.
- At the end of the GP, the band with EM similar to α -la was not visible indicating that this component was quite susceptible to proteolysis by pepsin in all four matrices [10]. However, clear differences in the gastric proteolysis of the band with EM similar to β -LG were observed among the four treatments. Namely, this protein remained pronounced, i.e., more pepsin resistant throughout the whole GP phase, in N-WPI compared to E-10 WPI, E-50 WPI, and H-WPI (*data not shown*).

A quite comparable proteolysis pattern was recorded by SDS-PAGE analysis (**Figure**). Based on the data provided by the current literature [3; 4; 11], the following observations are of interest:

- In GP, a fainter band corresponding to intact β -LG was evident in E-10 WPI, E-50 WPI, and H-WPI, compared to the native system (N-WPI).
- The greater degradation of β -LG in the above three denatured protein substrates (E-10 WPI, E-50 WPI, and H-WPI) was accompanied by the appearance of a set of lower molecular weight peptides (< 14.4 kDa). It should be highlighted that, under the conditions of this study, no such peptides were visualized in N-WPI.
- Regarding the IP phase, the degradation patterns of E-10 WPI and H-WPI exhibited many similarities as opposed to the degradation profile of E-50 WPI and N-WPI. At this point it should be mentioned, that according to Mao *et al.* (2019) [12], the presence of ethanol influenced not only the hydrolysis of intact β -LG but also the hydrolysate profiles, resulting from the action of trypsin.



Conclusions

Based on the preliminary data provided in this study, the structural modifications, induced by ethanol treatment, were shown to affect the release of the peptides generated during hydrolysis and consequently the digestibility of WP. Nevertheless, further investigation is required to confirm the aforementioned results and elucidate the underlying mechanisms of enzymic hydrolysis of ethanol-treated WP preparations.

References

1. Sanchón, J., *et al.* (2018). Protein degradation and peptide release from milk proteins in human jejunum. Comparison with *in vitro* gastrointestinal simulation. *Food chemistry*, 239, 486-494.
2. Hussein, F.A., *et al.* (2020). Whey Protein Concentrate as a Novel Source of Bifunctional Peptides with Angiotensin-I Converting Enzyme Inhibitory and Antioxidant Properties: RSM Study. *Foods*, 9(1), 64.
3. Nikolaidis, A., & Moschakis, T. (2018). On the reversibility of ethanol-induced whey protein denaturation. *Food Hydrocolloids*, 84, 389-395.
4. Feng, Y., *et al.* (2021). Ethanol induced changes in structural, morphological, and functional properties of whey proteins isolates: Influence of ethanol concentration. *Food Hydrocolloids*, 111, 106379.
5. ISO 1735. (2004). ISO 1735:2004 [IDF 5:2004], Cheese and processed cheese products - Determination of fat content - Gravimetric method (Reference method).
6. Brodkorb, A., *et al.* (2019). INFOGEST static *in vitro* simulation of gastrointestinal food digestion. *Nature protocols*, 14(4), 991-1014.
7. Minekus, M., *et al.* (2014). A standardised static *in vitro* digestion method suitable for food—an international consensus. *Food & function*, 5(6), 1113-1124.
8. Andrews, A.T. (1983). Proteinases in normal bovine milk and their action on caseins. *Journal of Dairy Research*, 50(1), 45-55.
9. Laemmli, U.K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680-685.
10. Cheison, S.C., & Kulozik, U. (2017). Impact of the environmental conditions and substrate pre-treatment on whey protein hydrolysis: A review. *Critical reviews in food science and nutrition*, 57(2), 418-453.
11. Li, S., *et al.* (2021). Impacts of heat-induced changes on milk protein digestibility: A review. *International Dairy Journal*, 105160.
12. Mao, Y., *et al.* (2019). β -Lactoglobulin hydrolysis by a flow-through monolithic immobilized trypsin reactor in ethanol/aqueous solvents. *Process Biochemistry*, 82, 84-93.