# Differences in the root anatomical traits of three *Salix* L. clones in response to increased Cd concentrations

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### INTRODUCTION

Among heavy metals, cadmium (Cd) is one of the most widely distributed pollutant, whose concentrations increase in the soil, water and air, globally and over time. Numerous physiological, biochemical and morphological studies have shown that woody species from the genus *Salix* L. (willows), represent suitable tools for phytoremediation of the sites polluted by Cd. However, only a few studies have followed structural changes of the root tissues exposed to increased concentrations of Cd in soil and water. Previous established studies physiological high Cd have accumulation in the roots of several willow clones, Salix alba L. clone 'B-44', Salix viminalis L. clone 'SV068', and Salix matsudana Koidz. clone 'SM404'. The aim of our work was to assess the effects of two single applied Cd concentrations (10<sup>-3</sup> and 10<sup>-6</sup> M Cd) on the roots' anatomical traits of these clones, grown in the soil contaminated with cadmium.

## **METHOD / DESIGN**

One-year old stem cuttings of three willow clones were obtained from the Institute for Lowland Forestry and Environment in Novi Sad, Republic of Serbia. The cuttings were grown in greenhouse of the Department of Biology and Ecology, Faculty of Science, Novi Sad, by the soil culture method. Our study included three treatments: control (without Cd), and treatments with 10<sup>-3</sup> M Cd and 10<sup>-6</sup> M Cd, each single added to the soil. Plants were harvested after 4 months of applied treatments (May to September). The anatomical response of the selected *Salix* clones to Cd was investigated on the microscopic sections of the part of the roots at a distance of 3 cm from the root neck. Examined roots developed secondary anatomical structure. Measurements of roots' cross-sections included following traits: root cross-section area, root diametar, thickness of the periderm, secondary phloem (cross-sectional area, percentage and thickness), secondary xylem (crosssectional area, percentage and diameter), cross-sectional area of parenchyma cells, and cross-sectional area, number and diameter of vessels.

Fig. 1. Root cross-sectional area of three *Salix* clones: A-C *Salix* alba L. clone 'B-44'; D-F Salix viminalis L. clone 'SV068'; G-I Salix matsudana Koidz. clone 'SM404' (A, D, G-control; B, E, Htreatment with 10<sup>-3</sup> M Cd and C, F, I-treatment with 10<sup>-6</sup> M Cd)

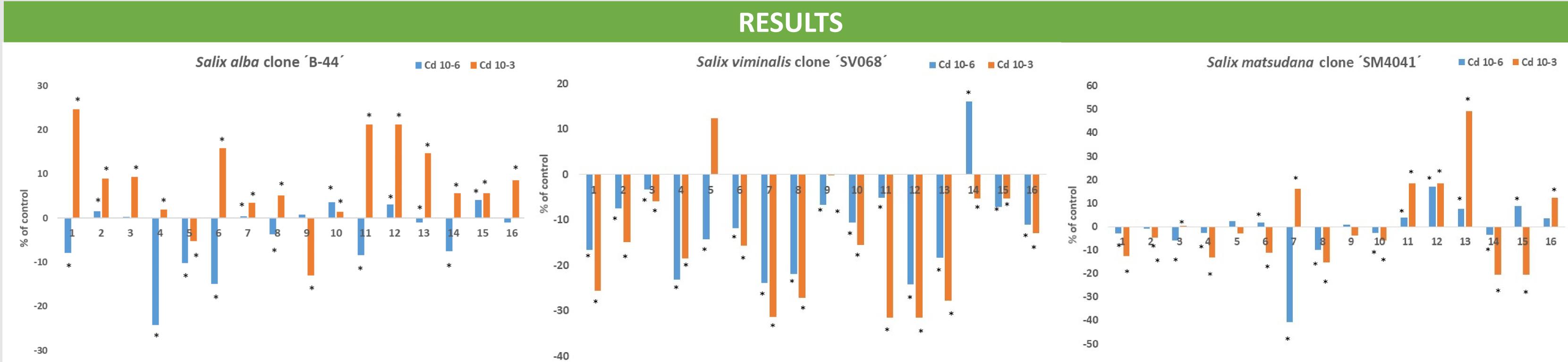


Fig. 2. Variations of measured roots' anatomical traits of Cd treated plants: 1 root cross-sectional area ( $\mu m^2 \times 10^5$ ); 2 root diameter ( $\mu m$ ); 3 thickness of the periderm ( $\mu m$ ); 4 crosssectional area of secondary phloem ( $\mu m^2 \times 10^5$ ); 5 percentage of secondary phloem; 6 thickness of secondary phloem; 7 thickness of secondary phloem; 6 thickness of secondary phloem; 7 thickness of secondary phloem; 6 thickness of secondary phloem; 7 thickness of secondary phloem; 7 thickness of secondary phloem; 8 thicknes; of secondary xylem (μm<sup>2</sup> x 10<sup>5</sup>); 9 percentage of secondary xylem; 10 diameter of secondary xylem; 11 total vessels area per treatment (μm<sup>2</sup> x 10<sup>4</sup>); 12 total vessel area per xylem cross-section (μm<sup>2</sup> x 10<sup>4</sup>); 13 cross-sectional area of vessels ( $\mu m^2$ ); 14 total vessels number per treatment; 15 vessels number per treatment; 15 vessels (Duncan test indicated significant differences between Cd-treated and control plants;  $*p \le 0.05$ )

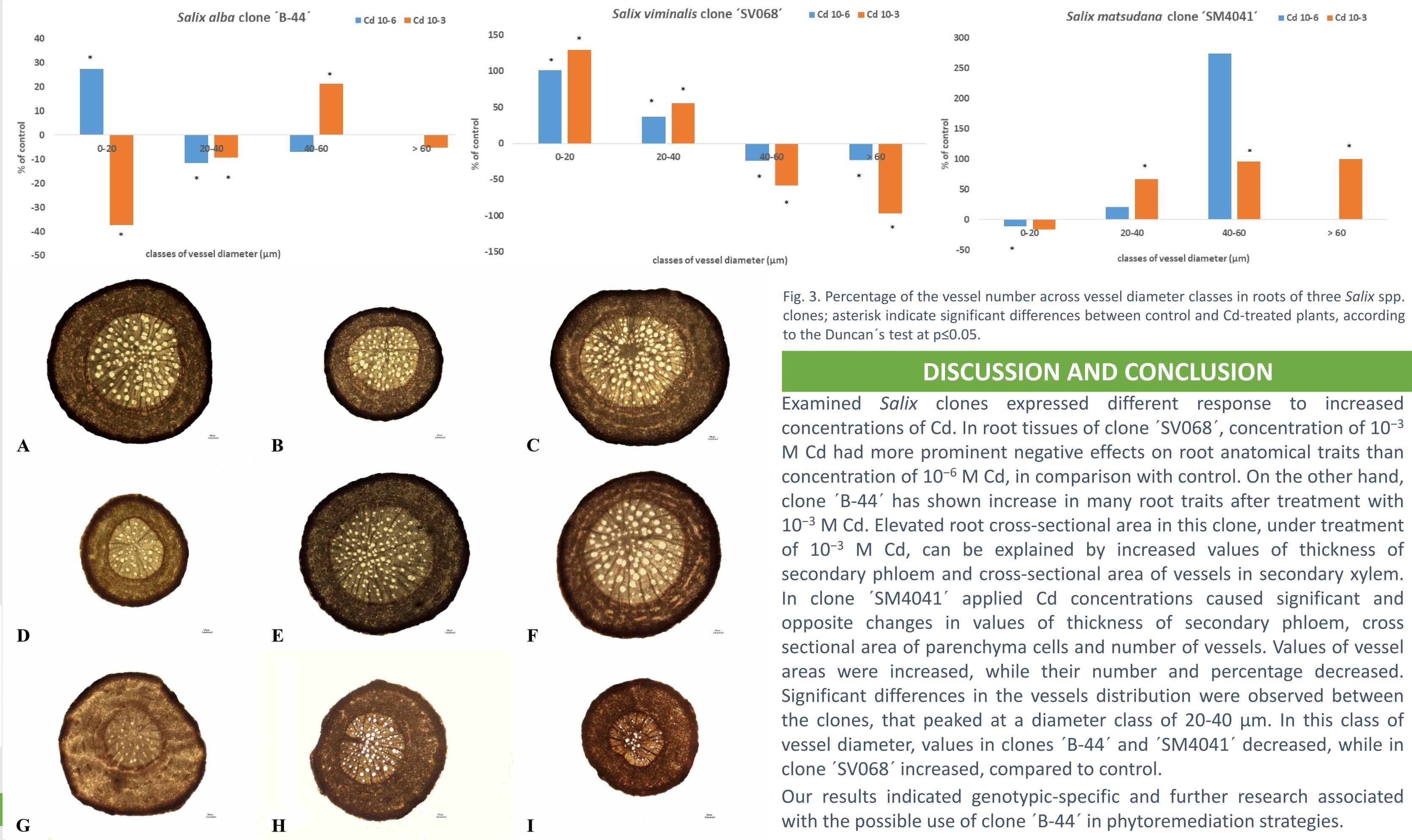


Fig. 3. Percentage of the vessel number across vessel diameter classes in roots of three Salix spp. clones; asterisk indicate significant differences between control and Cd-treated plants, according

