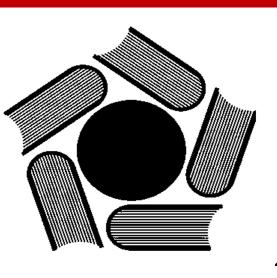
Defensive secretions of millipedes *Megaphyllum unilineatum* (C. L. Koch, 1838), Pachyiulus hungaricus (Karsch, 1881) and Cylindroiulus boleti (C. L. Koch, 1847) (Diplopoda, Julida) as antimicrobial agents in the inhibition of biofilms of **Pseudomonas aeruginosa PAO1 and Staphylococcus aureus**



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RESULIS

ANTIMICROBIAL ACTIVITY

IN RECENT YEARS: increased resistance of microorganisms to antibiotics

IMPERATIVE: discovery of new antimicrobial agents

- **BACTERIAL BIOFILMS** are highly resistant to antimicrobial agents which is serious global health concern
- plants and animals are a valuable source of natural biologically active products
- good basis for finding new antimicrobial and antibiofilm agents

OBJECTIVES

Determine the inhibition and degradation of the formed biofilm of *P. aeruginosa* PAO1 and *S. aureus* by defense secretions of selected millipede species from the family Julidae as well as to determine their antimicrobial activity by MTT assay.

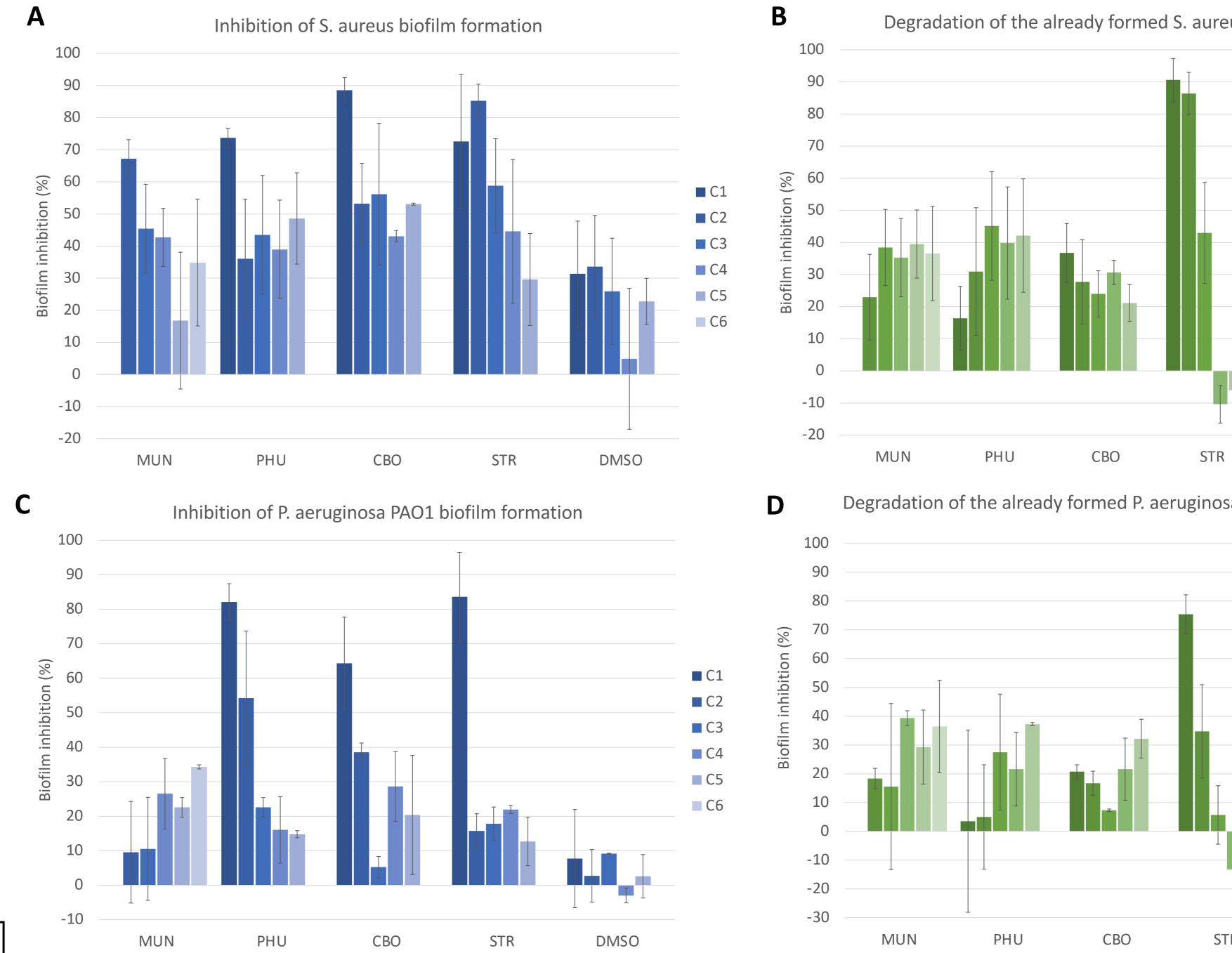
MILLIPEDES (DIPLOPODA)

- complex defense against predators
- various chemical compounds
- showed antimicrobial activity

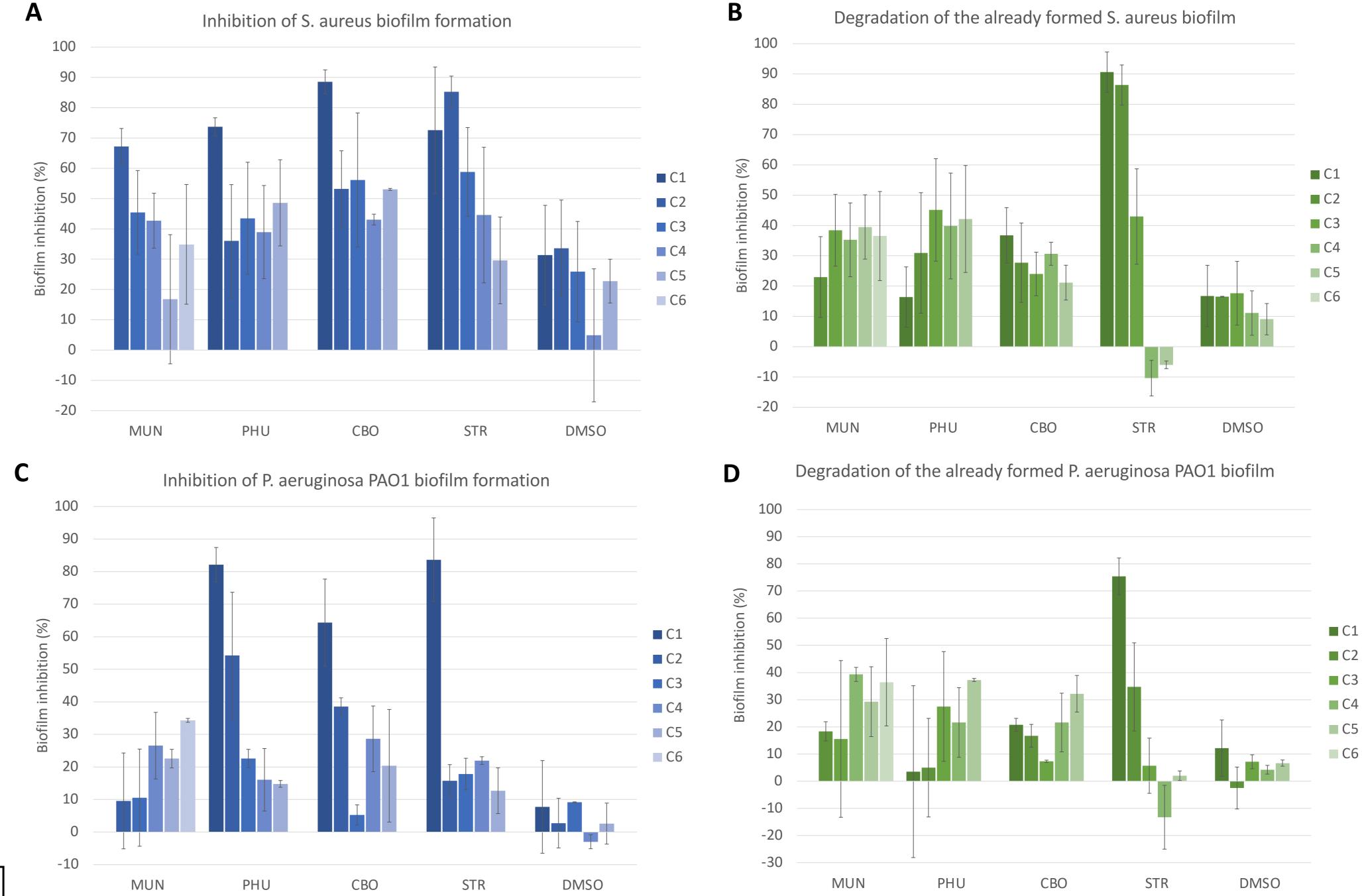
Defensive secretions showed a better antimicrobial activity against S. aureus compared to P. aeruginosa PAO1, which lacksquareis most likely due to the high resistance of this bacterial strain.

	S. aureus		P. aeruginosa (PAO1)	
mg/ml	MIC	MBC	MIC	MBC
MUN	0.03	0.06	>1	>1
PHU	0.06	0.12	1	>1
CBO	0.06	0.12	1	>1
Streptomycin	0.006	0.012	0.006	0.012
DMSO	>2.5%	>2.5%	>2.5%	>2.5%

ANTIBIOFILM ACTIVITY



Minimum inhibitory concentrations - MIC and Minimum bactericidal concentrations – MBC on S. aureus and P. aeruginosa PAO1 bacterial strains, presented in mg/mL.



ORDER JULIDA

Megaphyllum unilineatum (MUN) Pachyiulus hungaricus (PHU) Cylindroiulus boleti (CBO)

METHOD / DESIGN

BIOFILM FORMATION was quantified by the crystal violet staining method

ANTIMICROBIAL ACTIVITY was examined using the broth dilution minimum inhibitory concentration (MIC) test.

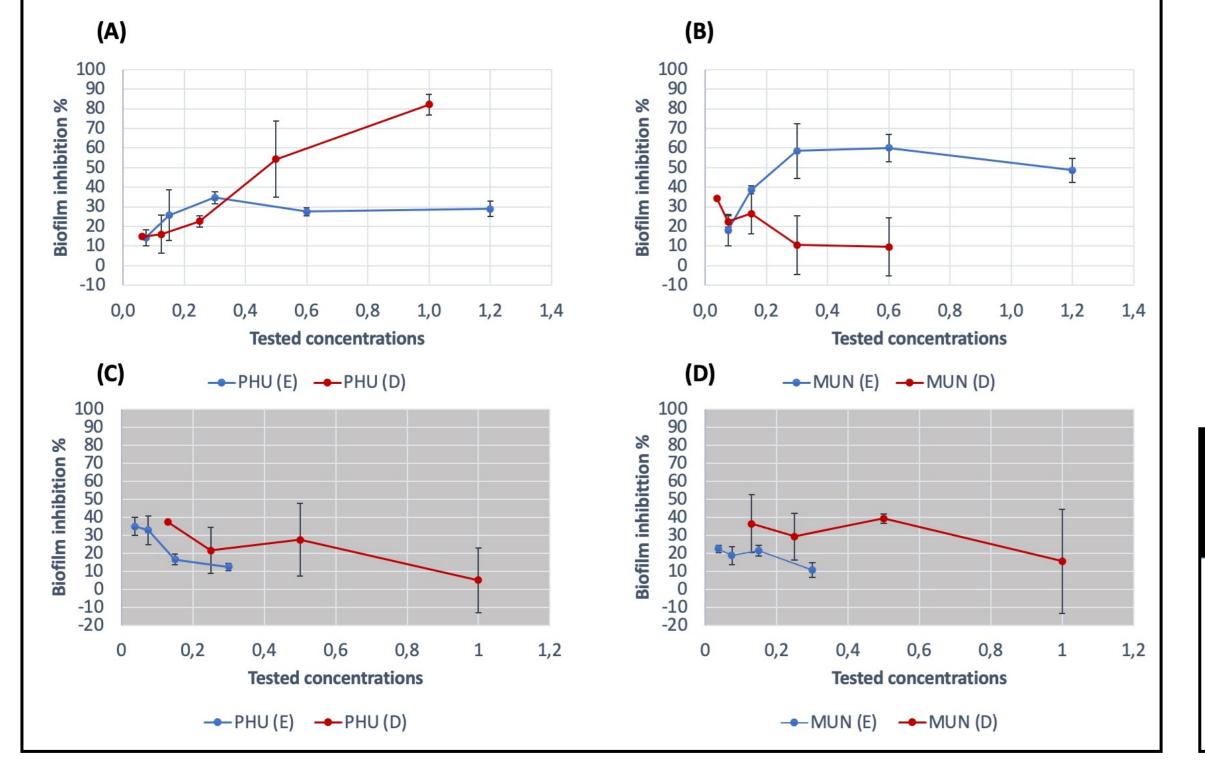
DMSO vs ETHANOL

- Inhibition of biofilm formation of defensive secretions (A) PHU and (B) MUN and degradation of the already formed *P. aeruginosa* PAO1 biofilm of (C) PHU and (D) MUN; both ethanol (E) and DMSO (D) extracts.
- DMSO extracts showed a stronger antibiofilm effect except in case

(A) Inhibition of S. aureus biofilm formation. Double dilutions were tested with the 2xMIC highest concentration tested (C1). (B) Degradation of the already formed S. aureus biofilm. Double dilutions were tested with the 4xMIC highest concentration tested (C1). (C) Inhibition of P. aeruginosa PAO1 biofilm formation. Double dilutions were tested with the highest concentration tested (C1) for MUN: 0.5 mg/ml, PHU and CBO: 1 mg/ml, DMSO: 1,25 % and streptomycin: 0,013 mg/ml. (D) Degradation of the already formed P. aeruginosa PAO1 biofilm. Double dilutions were tested with the highest concentration tested (C1) for MUN: 1 mg/ml, PHU and CBO: 2 mg/ml, DMSO: 2,5 % and streptomycin: 0,025 mg/ml.

• The antibiofilm effect was observed in all tested defense secretions and was more pronounced against S. aureus than against P. aeruginosa PAO1.

B, where ethanolic MUN extract showed a stronger effect in biofilm inhibition.



- The strongest biofilm inhibition of S. aureus was at the highest tested concentrations (2 × MIC) while degradation of already formed biofilm was shown at lower tested concentrations (MIC/4)
- The strongest inhibition of *P. aeroginosa* PAO1 biofilm formation was observed at the highest tested concentrations for PHU (82 and 54%), and CBO (64.3 and 38.5) while MUN had the strongest activity at the lowest tested concentration of 0.06 mg/mL (34.3%).
- All examined defense secretions had similar degradation activity of *P. aeruginosa* PAO1 biofilm with stronger activity at lower tested concentrations (about 30%).

CONCLUSIONS

The defense secretions of MBO, PHU, and CBO show a good basis for further investigations of their use as antimicrobial agents, especially against S. aureus.

Acknowledgements. This work was supported by the Serbian Ministry of Education, Science and Technological Development (Grant No. 451-03-9/2021-14/ 200178).