

Curriculum Vitae

Snezana Djordjevic

1. Appointment Details

Department: Biosciences - Structural and Molecular Biology

Present appointment: Professor in Structural Biology of Signalling and Head of Research Department of Structural and Molecular Biology

2. Education/Qualifications

10/2002	Accreditation of Teaching and Learning in Higher Education	UCL
01/1994	PhD (Biochemistry) "Structural Studies of Three Flavoproteins: NADPH-Cytochrome P-450 Reductase, Butyryl-CoA Dehydrogenase and Long Chain Acyl-CoA Dehydrogenase"	Medical College of Wisconsin, USA
06/1987	BSc Chemistry (Top of the class)	University of Nis, Yugoslavia

3. Professional History (in chronological order)

<u>Dates</u>	<u>Detail of position held</u>	<u>Institution</u>
07/2021 –	Head of Struct. and Mol. Biol.	UCL, London, UK.
08/2019 – 07/2021	Head of Struct. and Mol. Biol. (Interim)	UCL, London, UK.
10/2014 –	Professor	UCL, London, UK.
07/2006 – 07/2014	Senior Lecturer	UCL, London, UK.
12/1999 – 07/2006	Lecturer	UCL, London, UK.
11/1998 – 11/1999	Research Scientist	MRC LMB, Cambridge, UK.
11/1994 – 07/1998	HHMI Research Associate	Howard Hughes Medical Institute. Center for Advanced Biotechnology and Medicine (CABM), Piscataway NJ, USA.
01/1994 – 11/1994	Research Fellow	CABM, Piscataway NJ, USA.
04/1988 – 08/1988	Research Assistant	Medical College of Wisconsin, Milwaukee WI, USA.
09/1987 – 04/1988	Chemistry Teacher	Workers' University, Nis, Yugoslavia.

4. Other Appointments and Affiliations

10/2021 –	External Examiner, University of Oxford, Biochemistry
09/2019 –	Expert Assessor for the Science Fund of the Republic of Serbia, Belgrade, Serbia
02/2013 –	Visiting Professor in Biochemistry at the University of Novi Sad, Serbia
10/2013 – 03/2017	Consultant in Structural Biology and Drug Discovery for Magnus Life Science, UK.
03/2007 – 06/2012	Consultant in Structural Biology for Ark Therapeutics (links with Domainex Chemistry and BioFocus Crystallography units), UK.
08/1998 – 07/1999	Free-lance Editor for Macromolecular Structures, Elsevier, London, UK.

Professional Bodies

10/2013 –	Fellow, Royal Society of Biology
01/2007 –	Member of Biochemical Society
01/2007 –	Member of Biochemical Journal Editorial Advisory Panel
12/2006 – 01/2011	Committee Member of British Crystallographic Association; Biological Structures (Organizer of the BCA winter meeting 2007 together with Dr Parkinson UCL SOP)

Panel member Diamond Light Source, UK's national synchrotron science facility

Grant reviewer for: BBSRC, MRC, CRUK, Wellcome, Wellcome/DBT India Alliance, L'Oreal Women in Science

Peer reviewer for international standard journals in biological sciences including: Science Signalling, Biochemical Journal, Protein Science, PloS ONE, Journal of Molecular Biology, Acta Crystallographica, FEBS Journal.

5. Prizes, Awards and other Honours:

<u>Dates</u>	<u>Detail of prize, award or honour</u>	<u>Awarding/electing body</u>
2015	SLMS Teaching Award – team category (Course director)	
2013	Elected Fellow of the Royal Society of Biology, UK.	
1993	Friends of Medical College of Wisconsin Student Award	
1990	Scholarship for the Workshop on Molecular Mechanics and Dynamics of Biomacromolecules; Pittsburgh Supercomputing Center, Pittsburgh PA, USA.	

- 1988 Beatrice Kassell Award, Medical College of Wisconsin, Milwaukee WI, USA.
- 1988 Postgraduate Fellowship, Serbian Academy of Sciences, Belgrade, Yugoslavia.

6. Grants:

Industrial Funding and Private Investment:

Magnus Life Science

Title: 'Development of small molecule neuropilin antagonists', together with I Zachary, D Selwood, and P Frankel.

Date: 10/2013 – 08/2016.

Role: One of the four consultants within Magnus Life and Magnus Metabolic.

Description: Magnus Life and Magnus Metabolomics were formed in 2013, supported primarily by private funding. In the drug discovery programme focused on neuropilin-1, which initiated in 2014, Djordjevic provided key X-ray crystallography, structural biology and biophysics expertise supervising two post-doctoral scientists in the structural biology team. Generated > 30 ligand-bound structures.

ARK Therapeutics

Title: 'X-ray crystallography support for structure based development of neuropilin-1 antagonists'.

Date: 03/2007 – 03/2009.

Role: **PI**

Description: X-ray crystallography and structural biology support for Ark Therapeutics which included training and supervision of post-doctoral research assistant and contribution to the Research Committee.

Research Councils and Charities:

Wellcome Multi User Equipment Grant

Title: 'Strategic upgrade of crystal monitoring and optimisation equipment at ISMB, as critical infrastructure for structural biology and biochemistry'

Award Date: 2020

Award: £244,698

Role: **Lead UCL's PI**

BBSRC

Title: 'Clathrin assembly regulation of glucose metabolism'

Award Date: 2020; starting September 2021.

Award: £658,069 (requested, award letter pending)

Role: **Co-PI**

British Heart Foundation

Translational Award & Extension

Title: 'Lead optimisation of novel small molecule natriuretic receptor (NPR)-C agonists for the treatment of myocardial infarction'

Award Date: 2017-2020

Award: £524,524

Role: **Co-PI**

British Heart Foundation

Title: 'Structural and functional analysis of neuropilin-1 and neuropilin-2 interaction with VEGF'

Award Date: 02/2011-04/2014.

Award: £190,818

Role: **PI**

Medical Research Council (MRC)

Title: 'An integrated structural approach to a panel of pathogen-derived peroxidases with potential therapeutic targeting'

Award date: 09/2005-10/2008

Award: £218 860.

Role: **PI**

Biotechnology and Biological Sciences Research Council (BBSRC)

Title: 'Molecular basis of PP2A regulation through methylation-dependent assembly of heterotrimers'

Award Date: 03/2003 – 06/2006

Award: £176 000

Role: **PI**

BBSRC

Title: 'Rejuvenation of X-ray data collection facilities at the Institute of Structural and Molecular Biology'

Award Date: 2007

Award: £241,158

Role: **Co-PI** (Together with N Keep (PI), G Waksman, B Wallace, C Slingsby,...)

BBSRC

Title: 'High performance computing resources for structural biology and bioinformatics'

Award Date: 2005

Award: £179,940

Role: **Co-PI** (together with C Orengo (PI), P Driscoll, J Ladbury, S Perkins, D Jones)

BBSRC

Title: 'Computing resources for structural biology and bioinformatics'

Award Date: 2001-2004

Award: £308,000

Role: **Co-PI** (together with J Thornton (PI), P Driscoll, C Orengo and J Ladbury)

Internal Funding:

Research:

SLMS Capital Equipment Fund

Title: 'Upgrade of the Division of Biosciences Isothermal Calorimetry Instrumentation'

Award Date: 08/2018

Award: £ £86,000

Role: **PI**

SLMS Research Infrastructure Award

Title: 'Purchase of iTC200 and CD Spectrometer'

Award Date: 07/2012

Award: £79,000 (matched by £79,000 from the Division)

Role: **PI**

UCLB – Proof of Concept funding

Title: 'Expression, crystallisation and structural analysis of the human natriuretic peptide receptor (NPR)-C in the presence of novel receptor ligands'

Award Date: 2009 – 2010

Award: £50,000

Role: **Co-PI** (Together with A Hobbs (PI) and D Selwood)

Description: The project supported post-doctoral research assistant in Djordjevic's laboratory.

UCLH – Fast Track

Title: 'Design of expression constructs for NPRC'

Award Date: 2008 – 2009

Award: £30,000

Role: **Co-PI** (Together with A Hobbs (PI) and D Selwood)

Description: The project supported post-doctoral research assistant in Djordjevic's laboratory.

Teaching:

UCL Teaching Equipment Fund

Title: 'Purchase of new equipment to enable students to carry out protein gel and western blot analysis'

Award Date: 2012

Award: £1,886 (matched by £1,886 provided by the Division)

Role: **Lead Applicant**

Studentships:

Wellcome Trust PhD Studentship for Marta Wojnowska (2009-2012); ca £99,000
BBSRC PhD Studentship for Tamas Yelland (2012-2015); £73,778

7. Invited talks:

<u>Date</u>	<u>Details</u>
11/2021	Plenary Lecture, International Bioscience Conference, and the 8th International PSU-UNS Bioscience Conference IBSC 2020.
11/2019	Plenary Lecture, IX Conference of the Serbian Biochemical Society and a Poster Judge
03/2019	The 11 th National Forum on New Technologies in Drug Discovery, Shanghai, China.
11/2018	BioSense Institute, Novi Sad, Serbia
05/2018	St. Andrew's University, Scotland UK.
02/2018	Guest Lecture at the Trans-regional meeting at the TU University of Dresden, Germany.
10/2017	Department of Biochemistry University of Cambridge,
03/2017	The 9 th National Forum on New Technologies in Drug Discovery, Shanghai, China 2017
01/2016	Sensory Transduction in Microorganisms, Gordon Conference, Ventura CA, USA.
03/2015	The 7 th National Forum on New Technologies in Drug Discovery, Shanghai, China.
05/2015	Serbian Chemical Society, Novi Sad, Serbia.
05/2015	Inaugural Lecture, Division of Biosciences, UCL. London, UK.
02/2015	Sussex University, UK.
06/2014	COBRE Center in Structural Biology Annual Symposium, The University of Oklahoma, USA.
03/2013	The 5 th National Forum on New Technologies in Drug Discovery, Shanghai, (one of 25 invited speakers).
05/2012	London Structural Biology Club, London, UK.
10/2012	15 th International Symposium on Molecular Medicine, Crete, Greece, 2012 (unable to attend)
10/2011	25th Annual Centre for Advanced Biotechnology and Medicine (CABM) Symposium, Piscataway NJ, USA, 2011. (One of six CABM alumni selected to speak at the 25 th anniversary of the Centre)
10/2011	14 th International Symposium on Molecular Medicine, Rhodes, Greece.
08/2010	Drug Discovery session at the Queenstown Molecular Biology meeting, New Zealand.
08/2010	Tuberculosis session at the Queenstown Molecular Biology meeting, New Zealand.
08/2008	International Union of Crystallography meeting, iUCR Osaka, Japan.
07/2008	Computer Sciences Faculty, University of Nis, Serbia.

10/2008 CABM, Piscataway, NJ, USA.
10/2008 University of Sciences Pennsylvania, USA.
11/2007 National Institute for Medical Research, MRC, London, UK.
12/2007 Nucleic Acids Symposium, Prague, Check Republic.
05/2006 Institute of Child Health, London, UK.
10/2004 School of Biol. and Chem. Sciences, Birkbeck College, London, UK.
11/2003 London Structural Biology Club, London, UK.
06/2003 Bloomsbury Centre for Structural Biology Day, London, UK.
05/2001 Department of Crystallography, Birkbeck College, London, UK.
11/1999 London Structural Biology Club, London, UK.
01/1997 Bacterial Locomotion and Signal Transduction – BLAST, Quernavaca, Mexico, 1997

9. Research

PhD

- Structural biology of flavoenzymes. I made significant contribution to understanding of the substrate specificities of the fatty-acyl-CoA dehydrogenases. I crystallized and carried out initial crystallographic studies of a membrane-associated enzyme NADPH-Cytochrome P450 Reductase. Three major first author publications were generated as the result of that work.

Post-doctoral work

- Appointed as a Howard Hughes Medical Institute research associate at the Centre for Advanced Biotechnology and Medicine, Piscataway, NJ, USA. I described structural basis of receptor methylation-related regulation of bacterial chemotaxis. Specifically, I characterized methyltransferase CheR and the response regulator protein CheB which are the two enzymes involved in methylation/demethylation regulation of chemotaxis receptors (Djordjevic and Stock 1997; Djordjevic et al 1998; Djordjevic and Stock 1998). In addition, I structurally and functionally characterized interaction between CheR and the chemotaxis receptor Tar (Djordjevic and Stock 1998; Perez et al 2004).
- As an MRC-LMB research scientist (1998-1999) worked on the telomeric proteins and on the structure of yeast phenylalanine tRNA examining Mg²⁺ induced cleavage (Jovine et al 2000).

Upon arrival at UCL

- SD was recruited to UCL with a proposed research plan to structurally characterised methylation regulation of protein

phosphatase PP2A and the funding for this work was obtained in 2003. Meanwhile SD became an active member of the Bloomsbury Structural Biology Consortium and through the work-model of the consortium at the time she engaged in other collaborative work. This included collaborations with Prof Paul R Ortiz de Montellano (UCSF USA) on alkylhydroperoxidase AhpD from *M. tuberculosis*; with Prof J. Kelly and S Wilkinson at the London School of Hygiene and Tropical Medicine on novel peroxidases from *T. cruzi* and *T. brucei*; Prof J Ladbury on crystallisation of HNS oligomerisation domain. Through collaborations I entered the field of Cys-dependent peroxidases and managed to secure funding from MRC for some of this work (Nunn et al 2002, Koshkin et al 2003, Patel et al 2010).

Since 2005, structural biology of signalling

- At this point I decided to refocus my studies on signalling molecules. Following from my earlier work in bacterial chemotaxis I developed interest in two-component systems in *M. tuberculosis* and an autotrophic arsenite-oxidizing bacterium *Rhizobium sp. NT-26*. My laboratory was the first to show that *M. tuberculosis* histidine kinase DosS incorporates haem in order to support oxygen sensing and control of the dormancy regulon (Sardiwal et al., 2005). In collaboration with Dr J Santini, my lab has also characterized AioS/AioR system responsible for the control of arsenite oxidase genes in NT-26 (Sardiwal et al 2010; Wojnowska and Djordjevic 2012) and carried out identification and annotation of the two-component signalome of NT-26. SD's laboratory was engaged in structural and biophysical analysis and mechanistic description of a novel hexameric dual function histidine kinase/response regulator protein (Wojnowska et al 2013). My aim is to understand mechanistic properties of these modular signalling molecules and exploit that knowledge in the design of novel functional biological entities. Recently, we have solved the structures of the periplasmic arsenite-binding protein from NT-26 in the presence and in the absence of arsenite and characterised new class of oxyanion signalling proteins (Badilla et al 2018).
- Benefitting from the experiences gained in projects involving prokaryotic organisms and the knowledge of general principles of protein dynamics and protein chemistry, protein/protein interactions and mechanisms of signal propagation, SD laboratory is also actively involved in studying eukaryotic molecules involved in signalling.

- I am an Academic Lead for the ITC/CD spectrometry facility and the coordinator of the Darwin 6th floor communal laboratory space.
- In addition to academic collaborations, I participate in commercial translational medicine projects aimed at the development of small molecule antagonists of VEGF receptors neuropilin-1 and neuropilin-2. The work is underpinned by structural analysis of neuropilin's interaction with VEGF and other ligands, carried out in SD laboratory (Jarvis et al 2010; Seyedaraby et al 2013; Djordjevic and Driscoll 2013, Tsai et al 2016, Yelland et al 2016, Mota et al 2018). This is an exciting collaborative venture (Prof I Zachary, Prof D Selwood, Dr P Frankel) that integrates studies on neuropilin biology (previously funded by BHF project grant and BBSRC PhD Studentship) with studies of the small-molecule inhibitors in complexes with neuropilin receptors.
- I also have an established collaboration with Prof A Hobbs (formerly UCL, now Queen Mary University) and Prof D Selwood, working on the natriuretic peptide receptors.
- I am a co-applicant on a British Heart Foundation Translational Research Award, recently awarded to Hobbs, Selwood and Djordjevic for the development of NPRC agonists as a new therapy for cardiovascular diseases.
- Organizer of the ISMB external seminar series for Summer 2017, 'Signalling and beyond'.
- Developed collaborations with the research groups in Chemistry and Biomaterials at Free University of Berlin (Prof Rademann Jorg) and TU University at Dresden (Dr Vera Hintze), Germany which resulted in the exchange of materials, research visits and preparation of joint publication.

Technical Expertise

- I have strong expertise in various aspects of X-ray crystallography but I always used other structural, biochemical and biophysical techniques and have been engaged in collaborative research projects. X-ray crystallography studies have been, for example, complemented by NMR spectroscopy in collaboration with Prof Paul Driscoll (Parkinson et al 2008, Jarvis et al 2010, Patel et al 2010); molecular modelling (Djordjevic et al 1994, Perez et al 2004); calorimetry and SPR (Nunn et al 2005, Tsai et al 2010, Moyes et al 2014, Miyauchi et al 2016, Mota et al, 2018) or chemistry (Nunn et al 2002; Koshkin et al 2003, Jarvis et al 2011, Powell et al 2018). I have proven ability to integrate various methodologies and foster collaborations in order to answer the biological questions.

Academic supervision related

- Graduate Tutor for the research Department of Structural and Molecular Biology, since 09/2013.
- Co-Director for the WT PhD Programme
- ISMB Wellcome Trust PhD Programme Steering Committee 2013-2016
- SMB Representative on the Biosciences Graduate Staff-Student Consultative Committee
- Member of the FLS Research Degree Committee
- Member of the SMB committee for the assessment of student applicants for the Overseas PhD students scholarships.
- Member of Graduate School Funds sub-committee (2006 – 2013)
- Human Sciences Liaison Tutor (2001 – 2013)
- Interviewing Human Sciences UCAS candidates (2001 – 2012)

10. Knowledge Transfer

Consultancies

- X-ray crystallography/structural biology consultant for Ark Therapeutics, 2007-2012. Managed post-doctoral research assistant, participated at the Research Committee Meetings, Interacted with Chemistry section of Domainex and X-ray crystallography group of BioFocus, on behalf of Ark Therapeutics. Generated first crystal structure of the small-molecule antagonist and neuropilin-1 which initiated further crystallographic investigation of ligand/receptor complexes. Further 10 crystal structures determined either in Djordjevic or BioFocus laboratory.
- Consultant for Magnus Life Science. The two businesses were set up in continuation of the projects originally developed as part of Ark Therapeutics. Djordjevic was one of the four main scientists/consultants (together with I Zachary, D Selwood and P Frankel). More than 30 ligand-bound structures determined.
- Presented a short-talk at the UCLB-organized workshop on consultancies, 10/2013.

Academic exchange

- October 2021, External Examiner for the University of Oxford, Biochemistry.
- December 2017, External Examiner for the review of the new Oxford Biochemistry Degree Programme.

- Director, Continuing Professional Development Courses, Biosciences, UCL since 2011. Through the courses on 'Lab Techniques in Mammalian Cell Biology' and 'Antibodies: Sequence, Structure and Designing Therapeutics' we attracted participants from outside UCL and UK including, students, laboratory technicians, teachers, solicitors and industry scientists from Europe, USA and Africa. Obtained accreditation for the two courses by the Royal Society for Biology. Recently developed the Faculty of Life Sciences Policy document regarding the provision of short and CPD courses.
- 2015, SLMS Education Award –Teams Category for the work on CPD courses.
- At 2014, CALT conference, presented a talk 'From CPD courses to advanced final year research-based undergraduate modules'.
- In 2013, formally appointed Visiting Professor in Biochemistry at the Department of Chemistry, Biochemistry and Ecology, University of Novi Sad, Serbia. Developed two PhD-level one-week intense courses: 'Enterprise in life sciences' and 'Drug discovery process: from target validation to clinical trials'. The courses have passed accreditation and have become the part of the PhD programme since 2014.
- In February 2018 I was a lead academic from UCL on an Erasmus/Capacity Building application with the lead partners from Universities in Serbia and Lebanon and additional partners within FEBS, Universidade Nova de Lisboa, and World University Service, Austria.
- Involved in Yale/UCL Collaborative. Engaged in interaction with Prof J Schlessinger from Yale University regarding neuropilin-1/VEGF-R2 interaction.
- Visited Yale University as a part of the Yale/UCL Collaborative exchange, 2010
- Participated in two Dengue Fever workshops that included scientists from Yale, UCL and University of Queensland Australia.
- Involved in drug discovery project on NPR-C agonists with Prof D. Selwood at UCL and Prof A. Hobbs at Queen Mary University of London. Work supported by UCL Business 2009-2010.

Public engagement

- Enrolled in UCL's Inspiring Women in Science speakers programme.
- Since 2014 I gave several presentations at various high schools in London area.
- Participate in a 'Build My Future' conference in July 2016 in Bolton, UK.

- One of the four motivational speakers at the 2018 International Women's Day celebration at Plumstead Manor School, London, UK.

Professional bodies activities

- Organized (together with G Parkinson, UCL School of Pharmacy) British Crystallographic Association; Biological Structures Group Winter Meeting 2007.
- Member of the INSTRUCT HUB – Europe-wide scientific body that integrates infrastructure of expertise, technology platforms and education for furthering science.
- Contributed to the working group for writing VMX, a versatile micro-focus and in-situ diffraction facility for macromolecular crystallography proposal for DIAMOND Phase3.
- Participate in the INSTRUCT UK Structural Biology Forum meetings as ISMB representative
- Represented UCL at Rosalind Franklin Institute meetings.

PhD Viva examiner

- UCL degrees examinations: seven; University of London research degree examinations undertaken: ICR, Birkbeck, Queen Mary University, London School of Pharmacy, Crick Institute. External examiner at University of Oxford, Leicester University, and Sussex.

11. Teaching

Current Activities

- Module organizer for Bioc0008 Biomolecular Structure and Function since 2007. This is a core second year module, currently with nearly 120 enrolled students. The course includes lectures, five tutorials, six workshops and four practical sessions and Moodle page that I manage. I expanded curriculum, transformed the course from the 0.5 CU to the full CU, and organized additional workshops/case studies.
- Personal tutor to 4 students per each year of the degree programme (i.e. currently 12 students).
- Bioc0014 – Advanced Biomolecular Mechanisms, 6 hours
- Bioc0015 – The Molecular Basis of Cellular Regulation, 4 hours
- Bioc0003 – The Principles and Practice of Experimental Biochemistry, 10 hours (every other year)
- Bioc0008 – Biomolecular Structure and Function, 22 hours
- Bioc0021 – Literature review project, 4 students/year

- MSci students' supervision

Previous teaching engagement

- Teaching on MRes in systems biology, ABC Course, 4 hours.
- *Bioc3029 – Biosciences in Business and Media, 6 hours (I was a part of the working group designing this innovative module) (2010-2017)*
- Coordinator of the 'protein teaching stream' for Biochemistry/Mol. Biology Programme. Reviewing modules' contents to ensure flow of information and progression of curriculum through the years of the degree programmes. Running working groups at the Departmental 'Teaching away' days.
- Teaching on Biology of the Cell – specialized course for Human Sciences and Podiatry students, 2000-2006
- Course organizer for Biology of the Cell, 2001-2006. I reorganized course and exam structure and significantly improved pass rate of the Podiatry students on the course.
- Liaising with Podiatry programme and Board of Examiners 2001-2003.
- Human Sciences Liaison Tutor, 2001-2013. Active at the student-staff consultative committees, strategy and exam board meetings. Presented regularly at Human Sciences 'options days'. Interviewed UCAS candidates for Human Sciences degree programme.
- Member of the Organizing Committee for the Mathematics with Biology degree programme.
- Obtained Teaching Equipment Fund award (2012) for purchase of new horizontal SDS-PAGE systems and western-blotting equipment. New practical designed that would be carried out using the equipment.
- Course organizer for Bioc2007 Biochemistry for Biochemical Engineers as well as BengGB02, 2007-2011. These were students in Engineering (up to 50) that set on the same lectures as Bioc2004 but were not involved in practical sessions. I coordinated their exams and marks with the Engineering tutors and participated in the Engineering exam board meetings.
- MSc in Foetal genetics (2007 – 2012), 4 hours
- Bioc2004 – (2007 – 2011) Practical sessions, 12 hours
- B12 - Nucleic acids structure and function, replication, transcription and translation; Molecular biology and protein analysis methods (year 1)
- B100, General biochemistry tutorials (year 1)
- B11/B13 -- Haemoglobin structure and function, collagen and extracellular matrix (year 2)

12. Enabling

Management and organization

- Interim Co-Director for ISMB (since 2019)
- Biosciences Associate Director (Innovation and Enterprise, since 2019)
- CPD (Continuing Professional Development) Director for Biosciences, since 05/2011. Participated in CPD workshops organized by Prof J Dacre as a part of the UCL Health initiative. From 2011 focused on Biosciences. Generated business plan for the course 'Lab Techniques in Mammalian Cell Biology (I. Gout, academic lead)'. Piloted the course in 2012 with 14 students and managed organization of the course since 2013 with 20 paying participants and generating profits. Profits (in excess of £10,000/year) reinvested in purchase of new teaching equipment. In 2016, piloted new course: Antibodies: Sequence, Structure and Designing Therapeutics (A. Martin Academic lead). Obtained CPD accreditation for both courses through Royal Society of Biology. Liaising with Vice-Provost for Enterprise.
- Deputising for HoRD in variety of roles (2016-2019)
- Served on the promotions panel at the Divisional and SLMS level.
- Athena SWAN Academic Lead for the Division of Biosciences 2013-2016. Formed and managed self-assessment team for the Division. Chaired meetings, organized writing of the action plan, wrote first and second draft of the Bronze award application document. Bronze Award secured in 2015.
- Member of Biosciences SAT since 2016 working on a Silver Award application
- Member of the UCL Academic Board since 2006
- Co-organized the ISMB retreat in 2005
- Departmental seminar series organizer 2000-2005
- ISMB external seminar series organizer for the Summer of 2017

Equalities (in addition to Athena SWAN)

- Member of UCL Committee for Equal Opportunities (until 2013)
- Departmental Equal Opportunity Liaison Officer (until 2013)
- Chaired investigative panel for the Harassment and Bullying case (05/2012 – 04/2013)
- Member of the Disciplinary Appeal Hearing panel (06/2013)
- Presented a talk on 'Work/life balance' at the UCL Cell and Developmental Biology retreat in 2013
- Presented a talk 'University lecturer - master of many trades' at the ISMB retreat in 2007.

- Presented talks at Biology Society Retreats at Cumberland Lodge, Windsor, UK, January 2014 and January 2016.
- UCL Lunch Hour Lecture talk in January 2015: How can we deal with prejudice and gender bias in the collaborative and competitive world of science?

Recruitment

- Throughout the years participated in many recruitment panels for staff and post-doctoral scientists. For example, I was involved in the recruitment of: Prof Finn Werner, Prof Andrea Townsend-Nicholson, Dr John Kirkpatrick (UCL Facility Manager), Dr Nora Cronin (ISMB X-ray facility manager), Prof Christine Slingsby (Birkbeck), Dr Claire Bagneris (Birkbeck), Mr Mohamed Arooz (laboratory manager).
- 2018, on a recruitment panel for the new ISMB Director.

Other

- Participant in a 'Future Leader' Programme for 2015/2016
- Contributed to drafting Division of Biosciences Strategy for Enterprise at UCL 2011-2015.
- Academic representative for the Unified Email User Group assessing UCL transfer to the new email system 2009
- FLS Infrastructure review committee 2007
- Departmental computing committee (2002-2006)

SELECTED RECENT PUBLICATIONS

Conole D, Chou Y-T, Patsiarika A, Nwabo V, Dimitriou E, Soudy C, Mota F, Djordjevic, S., Selwood D. (2020). Discovery of a novel fluorescent chemical probe suitable for evaluation of neuropilin-1 binding of small molecules. *Drug Development Research*. doi:10.1002/ddr.21641.

Koehler, L., Ruiz-Gómez, G., Balamurugan, K., Rother, S., Freyse, J., et al., Djordjevic, S., Scharnweber, D., Rademann, J., Pisabarro M.T. & Hintze, V. (2019) Dual Action of Sulfated Hyaluronan on Angiogenic Processes in Relation to Vascular Endothelial Growth Factor-A. *9*, 18143. doi:10.1038/s41598-019-54211-0.

Badilla, C., Osborne, T.H., Cole, A., Watson, C., Djordjevic, S. & Santini, J. M. (2018). Arsenic oxyanion-binding proteins serve as sensors for regulating gene expression. *Scientific Reports* 8:6282 | doi:10.1038/s41598-018-24591-w

- Mota, F., Fotinou, C., Rhana, R., Chan, A., Yelland, T., Arooz, M. T., . . . Djordjevic, S. (2018). Architecture and hydration of the arginine binding site of Neuropilin-1. *FEBS Journal*. doi:10.1111/febs.14405
- Powell, J., Mota, F., Steadman, D., Soudy, C., Miyauchi, J. T., Crosby, S., . . . Djordjevic, S., Tsirka, S. E., Zachary, I. C., Selwood, D. L. (2018). Small molecule neuropilin-1 antagonists combine anti-angiogenic and anti-tumour activity with immune modulation through reduction of transforming growth factor beta (TGF β) production in regulatory T-cells. *Journal of medicinal chemistry*. doi:10.1021/acs.jmedchem.8b00210
- Yelland, T., & Djordjevic, S. (2016). Crystal structure of the neuropilin-1 MAM domain: completing the neuropilin-1 ectodomain picture. *Structure*, 24(11): 2008–2015. doi:10.1016/j.str.2016.08.017
- Miyauchi, J. T., Chen, D., Choi, M., Nissen, J. C., Shroyer, K. R., Djordjevic, S., Zachary, I. C., Selwood, D. & Tsirka, S. E. (2016). Ablation of neuropilin 1 from glioma-associated microglia and macrophages slows tumor progression. *Oncotarget*, 7(9), 9801-9814. doi:10.18632/oncotarget.6877
- Tsai, Y. C. I., Fotinou, C., Rana, R., Yelland, T., Frankel, P., Zachary, I., & Djordjevic, S. (2016). Structural studies of neuropilin-2 reveal a zinc ion binding site remote from the vascular endothelial growth factor binding pocket. *FEBS Journal*, 283(10), 1921-1934. doi:10.1111/febs.13711
- Windwarder, M., Yelland, T., Djordjevic, S., & Altmann, F. (2015). Detailed characterization of the O-linked glycosylation of the neuropilin-1 c/MAM-domain. *Glycoconjugate Journal*, 33(3), 387-397. doi:10.1007/s10719-015-9602-x
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